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Antioxidative Action of *Citrus limonum* Essential Oil on Skin

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Authors' contributions

The work presented here was carried out with the collaboration of all the authors. Authors GB and BT defined the research theme and designed the methods and experiments, analyzed the data, interpreted the results and wrote the paper. Author PA was involved in the writing process of the manuscript, author RV co-designed the experiments, discussed the analyses, interpretation, and presentation of data. All authors have contributed to, seen and approved the manuscript.

Research Article

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ABSTRACT

Aims: The purpose of this study was to investigate the action of *Citrus limonum* essential oil to control free radical-induced lipid peroxidation and preventing tissue damage in skin. **Place and Duration of Study:** Department of Internal Medicine (University of Roma "Tor Vergata) and A.R.P.A (Aging Research, Prevention and Therapy Association, www.antiaging.it), between January 2010 and June 2011.

Methodology: The essential oil was subjected to GC-MS analysis. The superoxide anion scavenging activity of essential oil was evaluated by the enzymatic hypoxanthine/xanthine oxidase system. The same oil diluted in DMSO or grape-seed oil was spread on the face of human volunteers after UV exposition. A sample of skin lipids was collected and the presence of peroxyl radicals was detected based on the measurement of light emitted (chemiluminescence) when the excited carbonyl and singlet oxygen decay to ground state.

Results: Our data demonstrate that the lemon essential oil is more active than α -tocopherol against O_2^- and peroxide free radical inhibition at 1:100 dilution. A protocol for controlling free radical-induced lipid peroxidation in human skin was thus proposed. **Conclusion:** The scavenging action of lemon essential oil could have a practical application for treating human skin against oxidative damage.

Keywords: Anti-aging; GC-MS; grape seed oil; superoxide anion scavenging.

1. INTRODUCTION

The inhibition of lipid oxidation by essential oils such as *Origanum* spp., *Thymus* spp., *Satureja* spp. and *Rosmarinus officinalis*, have already been reported in literature [1, 2, 3].

All the essential oils studied have shown a strong phenolic profile characterized by the presence of phenylpropanoids which are believed to be the active component of the essential oils [4, 5, 6, 7, 8, 9]. *Citrus* essential oil has also been reported to have antioxidative activities against linoleic acid oxidation [10] and to both Cu^{2+} -induced and 2, 2'-azobis (2-aminopropane) hydrochloride-induced oxidation of human low-density lipoprotein *in vitro* [11]. Among the compounds tested in *Citrus* essential oil, -terpinene had the strongest antioxidant effect [11], but no clear relationship could be shown between the antioxidant activity and the essential oil composition of the extracts [12]. When skin is exposed to air that is irradiated by ultraviolet (UV) light consisting of UVA (320-400 nm) and UVB (290-320 nm), reactive oxygen species (ROS) including superoxide anion radical (*O²⁻), hydrogen peroxide (H₂O₂), hydroxyl radical (*OH), singlet oxygen (*O₂), lipid peroxides (LOOH), and their radicals (LOO*) are formed. These in turn induce skin aging, phototoxicity, inflammation and malignant tumors [13, 14, 15, 16, 17].

Recently, consumer interest and the media have focused specifically on products that use natural ingredients, such as plant extract. There is some evidence that these ingredients could have possible *in vitro* anti-aging activity, but the question remains whether it is possible to deliver adequate doses to the skin *in vivo*. Lemon oil, traditionally used for its aromatic properties, has recently been investigated for its effects on skin [18]. The purpose of this study was to investigate the effectiveness of *Citrus limonum* Risso essential oil in controlling free radical-induced lipid peroxidation and preventing tissue damage in skin.

2. MATERIALS AND METHODS

2.1 Plant Material

The lemon (*Citrus limonum, Rutaceae*) essential oil used in this study was obtained from cold pressed oil extracted from the peel of the fruit (collected in the north of Sicily Island) according to the methods of Sawamura and Kuriyama [19]. The cold pressed oil was then hydrodistilled for 1h in an all-glass Clevenger apparatus. Voucher specimens of *C. limonum* plants, identified following the Italian botanical standard treatise [20] were deposited in the Herbarium of the Dept. of Applied Biology (University of Perugia, Italy).

2.2 GC and GC-MS Analysis

The GC analyses were carried out using a Varian 3300 instrument equipped with an FID and an HP-InnoWax capillary column (30 m x 0.25 mm, film thickness 0.17 μ m), working from 60°C (3 min) to 210°C (15 min) at 4°C/min or an HP-5 capillary column (30 m x 0.25 mm, film thickness 0.25 μ m) working from 60°C (3 min) to 300°C (15 min) at 4°C/min; the injector and detector temperature was 250°C. Helium was used as the carrier gas, with a flow rate of 1 ml/min, and the split ratio was 1:10.

GC-MS analyses were carried out with a Hewlett Packard 5890 GC-MS system operating in the EI mode at 70 eV, using the two above-mentioned columns. The operating conditions were analogous to those reported in the GC analyses section. The injector and transfer line temperatures were 220°C and 280°C, respectively. Helium was used as the carrier gas, with a flow rate of 1 ml/min and the split ratio was 1:10.

2.3 Identification of the Components

The components were identified by matching the spectra with those from mass spectral libraries; the identity of each component was confirmed by comparing the retention indices, from both columns, relative to the C6-C22 n-alkanes, with those from the literature [21, 22, 23, 24, 25]. When reported, co-elution gas chromatography with reference compounds was also used for an additional confirmation of the compound identity. The percentage composition of the essential oil was obtained by the normalization method from the GC peak areas, without using correction factors.

2.4 Superoxide Anion Scavenging (*O₂)

Superoxide anion was generated by a hypoxanthine-xanthine oxidase system [26]. Reaction mixtures with 100 μ I EDTA (30 mmol/l), 10 μ I hypoxanthine (30 mmol/l), 100 μ I cytochrome c (3 mmol/l) or nitroblue tetrazolium (3 mmol/l) were added to 150 μ I of lemon essential oil (solubilized in DMSO 10%) at various concentrations in a final volume of 3 ml buffered in KH₂PO₄ (50 mmol/l), pH 7.4 [27]. The reaction was started by adding 200 μ I xanthine oxidase (1U/ml) and the rate of reduced cytochrome c or nitroblue tetrazolium was measured at 550 nm, and 560 nm, respectively, against a reference. The amount of *O₂⁻ generated was calculated using the extinction coefficient ₅₅₀ = 2.1 x 10⁻² μ mol⁻¹ per cm and the *O₂⁻ inhibition was expressed as percentage values. The sample tested did not interfere with the xanthine oxidase activity (measured at 290 nm). The positive response was tested using -tocopherol. Ten repetitions were carried out.

2.5 Randomized Controlled Trial

2.5.1 Subjects

Subjects were selected from among men aged 18 to 52 (mean 33±11) years who were found to have no serious illness on physical checkup at A.R.P.A. (Aging Research, Prevention and Therapy Association), www.anti-aging.it (Civita Castellana, VT, Italy). Eighty volunteers (average age: 33±11 years) who gave their written consent to participate in the test were selected as subjects from January 2010 to June 2011.

2.5.2 Extraction of skin lipids from healthy volunteers

Skin lipids were collected with acetone-wetted cotton swabs from the forehead over a 9 cm² area from healthy volunteers (80 men, 18–52 years old–mean 33 ± 11) in the morning for 7 days. The sampling procedure was identical for all the subjects. The volunteers were randomly divided into four groups (A, B, C, D). In group A the forehead was treated daily for a week with -tocopherol in ethanol (20%), group B with lemon essential oil solubilized in DMSO (1:100), group C with lemon essential oil solubilized in grape-seed oil (1:100), and group D was left untreated. In accordance with the European norm EN 60335-2-27 and under medical supervision, volunteers were irradiated daily with UVA and UVB of 0.3 W/m^{2/}mm from sunlamps for 7 min at each session. The participants were asked not to expose themselves to direct sunlight and to avoid the use of face creams or hair lotions for the entire duration of the experiment. Twenty-four hours after the last treatment, the skin lipids were collected.

Extracts were taken twice from the wet cotton swabs using 3 ml of chloroform/methanol (1:2.5) for two hours (10 μ g heneicosanoic acid was used for the recovery test). The raw extracts were partitioned between 1% NaCl in 0.01 M HCl and chloroform. The chloroform extracts were washed with methanol/water (1:1) and dried under N₂ stream. The samples were stored at –20C^o in 3 ml of chloroform/ethanol (2:1).

2.6 Lipid Peroxidation Analyzed by Chemiluminescence

Chemiluminescence is an index of oxidative stress that quantifies lipid peroxidation and was measured according to the method of Gonzalez-Flecha et al. [28]. This method is based on the measurement of light emitted (chemiluminescence) when the excited carbonyl and singlet oxygen produced by peroxyl radicals decay to ground state. This light is due to the generation of reactive oxygen species in whole lipids. Skin lipids were incubated with 3 mM t-BHP for 60 min at 37°C. Lipid peroxidation was initiated by adding a small amount of stock solution of t-butyl hydroperoxide (80 mM) to each vial which was then maintained at 37°C, and measured by monitoring light emission [29] with a liquid scintillation analyzer Packard 1900 TR. Chemiluminescence was measured over a 60 min period and recorded as counts per minute (cpm) every 12 min. Each reaction was terminated by adding 5 ml chloroform/methanol (2:1, v/v) containing 0.01% butylated hydroxytoluene (BHT). This also inhibited any further oxidation during the lipid extraction. The DMSO had no antioxidative action and gave a chemiluminescence curve that could be superimposed on to that of the control.

2.7 Statistical Analysis

Analysis of variance, significances, correlations and other statistical analysis were performed using Graph Pad Prism version 5.00, (GraphPad Software, San Diego, California, USA).

3. RESULTS AND DISCUSSION

3.1 Chemical Composition of the Essential Oil

Citrus oils are a mixture of volatile compounds and consist mainly of monoterpene hydrocarbons [30]. *Citrus* essential oils, on the other hand, generally contain some amount of coumarins or furanocoumarins [31], flavonoids [32] and tocopherols [33] in the non-volatile

fractions of citrus oils. Coumarins and furanocoumarins may have an important role in skin photosensitization. Hydrodistillation of the cold pressed oil prevents this hazard. Nineteen compounds were identified in the GC and GC/MS analyses. The percentage composition of *Citrus limonum* essential oil is shown in Table 1. The components are listed in the order of elution from the HP-5 column. The main component was limonene (54.6%) followed by -terpinene (19.1%) and -pinene (14.5%). The monoterpene hydrocarbons (87.7%) constituted the main fraction of lemon oil. This oil composition, as reported in the literature, is similar to other volatile fractions characterized by the high content of limonene [34].

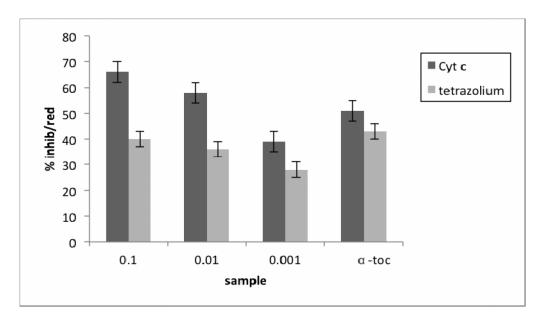
Compound	RI ^a	%
-pinene	938	3,9
-pinene	978	14,5
myrcene	993	1,5
-terpinene	1019	0,3
p-cymene	1024	0,1
limonene	1028	54,6
-terpinene	1061	19,1
terpinolene	1090	0,8
linalool	1098	0,1
citronellal	1154	0,1
terpinen-4-ol	1176	0,1
-terpineol	1189	0,3
citronellol	1225	0,1
nerol	1230	0,1
neral	1239	1,1
geraniol	1252	0,1
linalyl acetate	1258	0,1
geranial	1269	2,3
geranyl acetate	1383	0,8

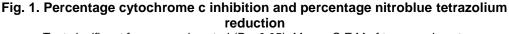
Table 1. Percentage composition of the essential oil from C. limonum

^aRetention index, relative to C_9 - C_{22} n-alkanes on the HP-5 column.

3.2 In Vitro and In Vivo Free Radical Scavenging Activity of Essential Oil

The superoxide anion scavenging activity of *Citrus limonum* essential oils was evaluated using the enzymatic hypoxanthine/xanthine oxidase system. Among the concentrations tested (Fig.1), the 1:100 dilution of lemon essential oil in DMSO had an $*O_2^-$ inhibition that was comparable to that of -tocopherol. The 1:1000 dilution inhibited $*O_2^-$ less than -tocopherol but the level of inhibition was about 76% and 65% of the -tocopherol activity on cytochrome c and tetrazolium nitroblue, respectively.





Test significant from normal control (P < 0.05). Mean <u>+</u>S.E.M of ten experiments

The peroxidation data as evidenced by the light emission are shown in Fig 2.

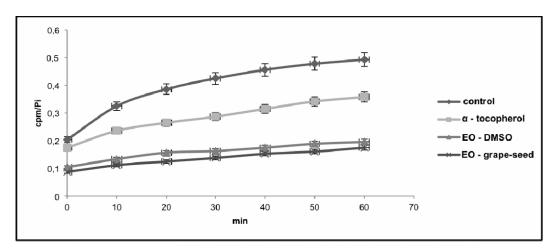


Fig. 2. Chemiluminescence over time in four groups of volunteers

r-tocopherol = group A; EO-DMSO = group B; EO-grape-seed = group C;control = group D. Test significant from normal control (P < 0.05). Mean \pm S.E.M of twenty experiments

The lipids from untreated volunteers showed the highest chemiluminescence and are considered to be the normal response to the peroxyl radical action. Lower emissions were recorded for the lipids from volunteers treated with antioxidative substances and the lemon essential oil was more effective than -tocopherol as an antioxidant. The grape-seed oil showed a slightly higher antioxidative action that was added to the action of lemon essential

oil; the chemiluminescence curve is a little lower than that of the lemon essential oil dissolved in DMSO, but the data belong to the same set according to the one-way ANOVA. These results show that these two oils had a similar scavenging action against peroxide free radicals *in vitro* and *in vivo* [35].

The exposure of human skin to UV radiation can generate ROS in both the epidermis and dermis. The depth of penetration of UV radiation, as well as its damaging potential in deeper skin cells have been demonstrated [36]. Among the scavenging substances, -tocopherol was chosen as a reference for comparing the scavenging action of lemon essential oil. The anti-oxidant activity of oil-in-water emulsion containing -tocopherol has been reported over a wide range of conditions and test systems [37]. Our data demonstrate that the lemon essential oil is more active than -tocopherol against $*O_2$ and peroxide free-radical inhibition at 1:100 dilution. Lemon essential oil is used instead of other lemon extracts, to avoid the toxic action that furanocoumarins have under UV exposure.

4. CONCLUSIONS

The results of this study suggest that lemon essential oil has properties that could benefit human skin as it undergoes environmental and chronological ageing. The scavenging action of lemon essential oil solubilized in grape-seed oil could have a practical application in aesthetic medicine (a branch of medicine focused on satisfying the aesthetic desires and goals of patients) for treating human skin against oxidative damage. Therefore, continuous application of lemon essential oil solubilized in grape-seed oil might contribute to the prevention of lifestyle-related skin diseases by regulating the balance of oxidative stress.

ETHICAL APPROVAL

All authors hereby declare that all experiments have been examined and approved by the appropriate ethics committee and have therefore been performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki.

COMPETING INTERESTS

The authors declare that no competing interests exist.

REFERENCES

- 1. Estevez M, Cava R. Effectiveness of rosemary essential oil as an inhibitor of lipid and protein oxidation: contradictory effects in different types of frankfurters. Meat Sci. 2006;72(2):348-355.
- 2. Kulisic T, Radonic A, Milos M. Inhibition of lard oxidation by fractions of different essential oils. Grasas y Aceites (Sevilla, Spain). 2005;56(4):284-291.
- 3. Nakatsu T, Lupo AT, Chinn JW, Kang RKL. Biological activity of essential oils and their constituents. Stud Nat Prod Chem. 2000;21(B):571-631.
- 4. Teissedre PL, Waterhouse AL. Inhibition of oxidation of human low-density lipoproteins by phenolic substances in different essential oils varieties. J Agric Food Chem. 2000;48(9):3801-3805.
- 5. Angelini P, Pagiotti R, Menghini A, Vianello B. A antimicrobial activities of various essential oils against foodborne pathogenic or spoilage moulds. Ann Microb. 2006;56(1):65-69.

- 6. Angelini P, Granetti B, Pagiotti R. Effect of antimicrobial activity of *Melaleuca alternifolia* essential oil on antagonistic potential of *Pleurotus* species against *Trichoderma harzianum* in dual culture. World J Microb Biotech. 2008;24(2):197-202.
- 7. Angelini P, Pagiotti R, Venanzoni R, Granetti B. Antifungal and allelopathic effects of Asafoetida against *Trichoderma harzianum* and *Pleurotus* spp. Allelopathy J. 2009;23(2):357-368.
- 8. Pagiotti R, Angelini P, Rubini A, Tirillini B, Granetti B, Venanzoni R. Identification and characterisation of human pathogenic filamentous fungi and susceptibility to *Thymus schimperi* essential oil. Mycoses 2011;54(5):e364-76.
- 9. Tirillini B, Pagiotti R, Angelini P, Pintore G, Chessa M, Menghini L. Chemical composition and fungicidal activity of the essential oil of *Laserpitium garganicum* from Italy. Chem Nat Comp. 2009;45(1):103-105.
- 10. Song H-S, Ukeda H, Sawamura M. Antioxidative activities of citrus peel essential oils and their components against linoleic acid oxidation. Food Sci Technol Res. 2001;7(1):50-56.
- 11. Takahashi Y, Inaba N, Kuwahara S, Kuki W. Antioxidative effect of citrus essential oil components on human low-density lipoprotein *in vitro*. Biosc Biotech Biochem. 2003;67(1):195-197.
- 12. Di Vaio C, Graziani G, Gaspari A, Scaglione G, Nocerino S, Ritieni A. Essential oils content and antioxidant properties of peel ethanol extract in 18 lemon cultivars. Sci Hortic. 2010;126(1):50–55.
- 13. Bech-Thomsen N, Wulf HC. Carcinogenic and melanogenic effects of a filtered metal halide UVA source and a tubular fluorescent UVA tanning source with or without additional solar-simulated UV radiation in hairless mice. Photochem Photobiol. 1995;62(4):773-779.
- 14. Kligman AM. Early destructive effect of sunlight on human skin. J Am Med Assoc. 1969;210(13):2377-2380.
- 15. Oikarinen A, Karvonen J, Uitto J, Hannuksela M. Connective tissue alterations in skin exposed to natural and therapeutic UV-radiation. Photoderm J. 1985;2(1):15-26.
- 16. Sakurai H, Yasui H, Yamada Y, Nishimura H, Shigemoto M. Detection of reactive oxygen species in the skin of live mice and rats exposed to UVA light: a research review on chemiluminescence and trials for UVA protection. Photochem Photobiol Sci. 2005;4(9):715-720.
- 17. Watson REB, Griffiths CEM. Pathogenic aspects of cutaneous photoaging. J Cosm Dermat. 2005;4(4):230-236.
- Chiu A, Kimball AB. Topical vitamins, minerals and botanical ingredients as modulators of environmental and chronological skin damage. Br J Dermatol. 2003;149(4):681-691.
- 19. Sawamura M, Kuriyama T. Quantitative determination of volatile constituents in the pummelo (*Citrus grandis* Osbeck forma Tosa-buntan). J Agric Food Chem. 1988;36(3):567–569.
- 20. Pignatti S. Flora d'Italia. Edizioni Edagricole, Bologna, Italy; 1982.
- 21. Adams RP. Identification of essential oil components by gas chromatography/quadrupole mass spectroscopy. Allured Publishing Corporation; 2001.
- 22. Davies NW. Gas chromatographic retention indices of monoterpenes and sesquiterpenes on methyl silicone and Carbowax 20M phases. Chromatogr. 1990;503:1-24.
- 23. Heller SR, Milne GWA. EPA/NIH mass spectral data base U. S. Government Printing Office; 1983.

- 24. Jennings WG, Shibamoto T. Qualitative analysis of flavour and fragrance volatiles by glass capillary gas chromatography. Academic Press; 1980.
- 25. McLafferty FW, Staufer DB. The Wiley NBS registry of mass spectral data. John Wiley and Sons; 1989.
- 26. Aruoma O, Halliwell B, Hoey BM, Butler J. The antioxidant action of N-acetylcysteine: its reaction with H₂O₂, OH^{*}, OH^{*}₂, HOCI. FRBM. 1989;6(6):593-597.
- Gressier B, Cabanis A, Lebegue S, Brunet C, Dine T, Luyckx M, Cazin M, Cazin JC. Decrease of hypochlorous acid and hydroxyl radical generated by stimulated human neutrophils: comparison *in vitro* of some thiol-containing drugs. Methods Find Exp Clin Pharmacol. 1994;16(1):9-13.
- Gonzalez-Flecha B, Llessuy SF, Boveris A. Hydroperoxide-initiated chemiluminescence: an assay for oxidative stress in biopsies of heart, liver and muscle. Free Radic Biol Med. 1991;10(2):93–100.
- 29. Wright JR, Rumbaugh RC, Colby HD, Miles PR. The relationship between chemiluminescence and lipid peroxidation in rat hepatic microsomes. Arch Biochem Biophys. 1979;192(2):344-351.
- Dugo P, Mondello L, Cogliandro E, Cavazza A, Dugo G. On the genuineness of citrus essential oils. Part LIII. Determination of the composition of the oxygen heterocyclic fraction of lemon essential oils (*Citrus limon* (L.) Burm. f.) by normal-phase high performance liquid chromatography. Flavour Fragr J. 1998;13(5):329-334.
- 31. Dellacassa E, Lorenzo D, Moyna P, Verzera A, Mondello L, Dugo P. Uruguayan essential oils. Part VI. Composition of lemon oil. Flavour Fragr J. 1997;12(4):247-255.
- 32. Miyake Y, Yamamoto K, Osawa T. Isolation of eriocitrin(eriodictyol 7-rutinoside) from lemon fruit (*Citrus limon* Burm. f.) and its antioxidative activity. Food Sci Technol Int Tokyo. 1997;3(1): 84-89.
- 33. Waters RD, Kesterson JW, Braddock RJ. Method for determining the -tocopherol content of citrus essential oils. J Food Sci. 1976;41(2):370-371.
- Espina L, Somolinos M, Lorán S, Conchello P, Pagán DGR. Chemical composition of commercial citrus fruit essential oils and evaluation of their antimicrobial activity acting alone or in combined processes. Food Control. 2011;22:896-902.
- 35. Ahn HS, Jeon TI, Lee JY, Hwang SG, Lim Y, Park DK. Antioxidative activity of persimmon and grape seed extract: *in vitro* and *in vivo*. Nutr Res. 2002;22(6):1265-1273.
- Katiyar SK, Afaq F, Perez A, Mukhtar H. Green tea polyphenol (-)-epigallocatechin-3gallate treatment of human skin inhibits ultraviolet radiation-induced oxidative stress. J Carcinog. 2001;22(2):287-294.
- 37. Frankel EN, Huang SW, Kanner J, German JB. Interfacial phenomena in the evaluation of antioxidants: bulk oils vs emulsions. J Agric Food Chem. 1994;42(5):1054-1059.

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