



The Importance of Isolated and Purified *Bacillus* species as a Source of a Variety of Antimicrobial Substances

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Author's contribution

The sole author designed, analysed, interpreted and prepared the manuscript.

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ABSTRACT

The rapid onset of resistance reduces the efficacy of most conventional antimicrobial drugs and is a general cause of concern for human well-being. Thus, there is a great demand for a continuous supply of novel antibiotics to combat this problem. Bacteria-derived antimicrobial peptides (AMPs) have long been used as food preservatives. Thus, antimicrobial agents are predominantly significant in limiting infectious diseases. Meanwhile, the development and dissemination of drug-resistant strains in pathogenic bacteria have begun to be a significant general health threat. Therefore, the lowest effectiveness of the present antibiotics in the direction of the administration was becoming prominent to the consequence of the discussion of new antibiotics. These considerations, *Bacillus* sp completely famous for making different metabolites of the possibility their utilization, thus, it is completed a search to isolate *Bacillus* sp for the action making of antimicroorganisms compounds. The Biosynthesis of *Bacillus* AMPs, extract, purifies, and identifies a bioactive compound by *Bacillus* sp. Furthermore, *Bacillus*-derived AMPs can be synthesized both ribosomal and non-ribosomal and can be classified according to peptide biosynthesis, structure, and molecular weight. The precise mechanism of action of these AMPs is not yet clear; however, the one proposed mechanism is that

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these AMPs kill bacteria by forming channels in and (or) disrupting the bacterial cell wall. *Bacillus*-derived AMPs have potential in the pharmaceutical industry, as well as the food and agricultural sectors. Here, we focus on *Bacillus*-derived AMPs as a novel alternative approach to antibacterial drug development.

Keywords: *Bacillus sp.*; stimulation; antimicrobial compound.

1. INTRODUCTION

The fast start of resistance decreases the effectiveness of great traditional anti-microorganisms drugs. Therefore, there is large request for a continued make available new antibiotics to treat the trouble. Antimicrobial peptides (AMPs) were utilized as a protection to materials for foods. At a recent time, peptides from the genus *Bacillus* had contained a spectrum of anti-microorganisms activity against diseases microorganism. Antimicrobial peptides (AMPs) were produced from each ribosomal and non-ribosomal using peptide biosynthesis and it was suggested that these AMPs kill bacteria with damage to the bacterial cell wall. AMPs as considerable new more available possibilities come near to antibacterial drug advancement and supply a summary of a subject of the mode of action, implementation, and efficiency various AMPs make by members of the *Bacillus* genus containing new AMPs [1].

This review article was to illustrate the *Bacillus* species, for assurance on novel results which emphasize their more available possibilities to traditional antibiotics.

1.1 Biosynthesis *Bacillus* AMPs

Antimicrobial peptides (AMPs) had consisted of amino acids and made by all classes of multicellular organisms as an absolutely necessary of the immune response. After that the membrane damage happens with different mechanisms, driving to damage its safety also in the end, cell death [2]. AMPs apply intracellular inhibitory activity by interfering with different absolutely necessary processes. Moreover, in supplement to anti microorganism activities, also AMPs adjust the immune response activating cytokine production, and happening injury healing [3].

Antimicrobial peptides (AMPs) had contained the possibility to play showing signs of future success to combat against the elevate in microorganisms' resistance of traditional antibiotics Rotem and Mor 2009. Many various

AMPs produced from plants, animals and microorganisms also it more was entered into a clinical experiment Brogden and Brogden [4]. The AMPs are a defense system for human anti microorganisms', undesirable side impacts are less probably than with chemical antibiotics. Furthermore, the AMPs are about carefully to be a primary group of new anti-microorganisms drugs utilized in the therapy of parasitic and infectious diseases, it could appropriate to the therapy of chronic diseases [4]. Traditional antibiotics may selectively improve the resistance, meanwhile, AMPs kill microorganisms firstly during the membrane pores, and therefore work is essentially a great hardship to promote resistance [5]. *Bacillus* is competent in making AMPs, thus considered as a favorable in as novel inhibitory substances [6]. More research has found that the genus *Bacillus* sensuality make a more anti microorganisms' substance had contained lipopeptides, bacteriocins, and other kinds of peptides [7].

A recent study of the synergistic effect between a broad set of AMPs and antibiotics like ciprofloxacin, meropenem, erythromycin, and vancomycin for treating infections caused by clinical hard-to-treat pathogens, including all ESKAPE (*Enterococcus faecium*, *S. aureus*, *K. pneumoniae*, *A. baumannii*, *P. aeruginosa*, *Enterobacter cloacae*) pathogens, revealed the ability of AMPs to elicit degradation of ppGpp, avoiding the entry in an energy-starved state. This is a significant finding as potentially all microorganisms react to antibiotic treatment mediating persistence, and the fact that some AMPs can prevent it opens new doors for the development of alternative therapies that effectively decrease the resistance rate [8].

Members of the genus *Bacillus* are rod-shaped, endosporeforming, Gram-positive bacteria that are abundant in soil. The *Bacillus* species can produce structurally diverse secondary metabolites, which exhibit a wide spectrum of antibiotic activity [9]. The peptide antibiotics of the *Bacillus* species can be divided into 2 subgroups based on the synthesis pathway [10]. One of these subgroups includes small microbial peptides that are non-ribosomal synthesized by

large enzymatic complexes, whereas the second subgroup comprises ribosomal synthesized peptides [11].

1.2 Purified/ Extract Antimicrobial Activity from Media

More strains of *B. amyloliquefaciens* are recognized as anti-microorganisms [12]. Recently, Torres et al. (2013) reported that the antiviral activity for peptide and also, the N3 lipopeptide, made from *B. amyloliquefaciens* M1 strain, it showed that anti microorganisms' activity. This lipopeptide had contained amino acid and hydroxy fatty acids. Therapy with the lipopeptide N3 was appeared to consequence in damage and kills the bacterial cell [13].

Bacillus mojavensis A21 has produced the non-ribosomal made by synthesis lipopeptide biosurfactants which considerable activity against anti microorganisms [14]. Chopra et al. [15] specified new AMP namely sonorensin produced from *Bacillus sonorensis* MT93. Furthermore, AMP is exhibited antimicrobial activity against popular foodborne pathogens. Moreover, the extracts from the cell-free of 2 *Bacillus* species, had could exhibit strong as an anti-microorganisms activity against phytopathogens in experiments and it recommended for the biosynthesis of the lipopeptides [16].

At last years, the reduced effectiveness of present antibiotics directed the administration of becoming prominent drug-resistant strains had required studying to new antibiotics. *Bacillus sp* is potential use to making a difference of minor metabolites. Thus, it could be carried out to isolate *Bacillus sp* from the oral cavity for the production of new antimicrobial compounds, then identified a new bioactive compound by *B. megaterium*. The bioactive compound was extracted by using N-butanol purified and known as a cyclic polypeptide and its similarity with bacitracin [17].

1.3 The Useful Functions to AMPs of the Bacillus Species

The *Bacillus sp* is significant due to their fast growth ratio, which consequences in low fermentation cycles; and it able for protein excretion into the situated or taking place outside a cell medium [12]. More *Bacillus* species were utilized as a probiotic complement in both animals and humans [18].

The polypeptides are the greatest importance of bacitracin, therefore it utilized in with anti microorganism agents [19]. For this reason, the cyclic and anionic AMP subtilosin, has been explained to apply as an anti microorganism activity against human pathogens [20]. Polymyxin B and E is fast bactericidal AMPs and also it is anti- microorganisms had contained Enterobacteriaceae and non-fermentative species [21]. For the reason the danger side influences, at the present time, it has been utilized as a cream for surface wounds [22].

At a recent time, antimicrobial lipopeptide microcapsules were examined as food preserves it [23]. Subtilosin is substance food preservation in order of its effectiveness against microorganisms and foodborne diseases [24]. Amylolysin, a new bacteriocin, which is food-related listeriosis and inhibition of the various *L. monocytogenes* isolates in chickens meat. Therefore, it utilized to protective in chickens meat [25].

Lipopeptides like surfactin has emulsification characteristics indicating to AMPs have a usage in biotherapy. The surfactin lipopeptide observed that as antitumor, ant-microorganisms, and lowering cholesterol [26]. More *Bacillus*-species-derived AMPs and *B. subtilis* species can be used to inhibit plant pathogens, biocontrol of plant diseases and safeguard grain [26].

2. EXTRACTION AND PURIFICATION γ -ACTINORHODIN

γ -actinorhodin possesses more necessary characteristics as anti microorganisms drug, proving strong an against key Gram-positive pathogens. Results confirmed the advantage not used to benefit antibiotics as a source of new antibacterial drug [27].

Actinorhodin (ACT), is an antibiotic made by *Streptomyces coelicolor* Silver [28]. It was afterwards observed that *S. coelicolor* make from a collection of compounds which are as a whole indicated to actinorhodins (ACTs). More research appears that used mixtures of ACTs, for the technique utilized properties ACT are not enough to dissolve it from its close analogues [29].

3. ISOLATION AND PURIFICATION OF γ -ACTINORHODIN (γ -ACT)

Less knowledge occurs concerning the characteristics of anti-microorganisms. It found

low antibacterial activity against some Gram-positive bacteria. Moreover, the ACT its antibacterial influence by catalysing the action of making toxic levels of hydrogen peroxide. Preparatory detected more pigments displaying the litmus-like characteristics of ACTs; which known it as γ -ACT (Fig. 1a). Thereafter, it was isolation and purification of this compound (Fig. 1b) [27].

4. IN VITRO ANTIBACTERIAL ACTIVITY OF γ -ACT

γ -ACT is against Gram-positive diseases bacteria (Fig. 2a). The antistaphylococcal activity of γ -ACT was determined against a panel of 70 clinical isolates of *S. aureus* methicillin-resistant (Fig. 2b). Therefore, the antistaphylococcal potency of γ -ACT did not show activity against Gram-negative pathogens (Fig. 2) may be the reason that they are unable to through the outer membrane (OM) [30]. Meanwhile, no alteration in

antibacterial activity of γ -ACT was showed versus strains influence deleted.

5. IDENTIFICATION OF THE ACTIVE COMPOUND

Antimicrobial peptides AMPs are found for many forms bacteria isolated from plants, and animals [31] and to give care against microorganisms and are made by the synthesis in fat bodies and in blood cells [32]. It can have local synthesis in various tissues, like the gut and epidermis [33].

A new antimicrobial peptide, named Bicarinalin, was isolated from the venom of the ant. It had a strong and anti microorganisms activity of the same strength as Melittin. Furthermore, this anti microorganisms' peptide has a low hemolytic activity than to Melittin on erythrocytes, point out the possibility for development into an anti-infective agent for utilizing against emerging antibiotic-resistant pathogens [34].

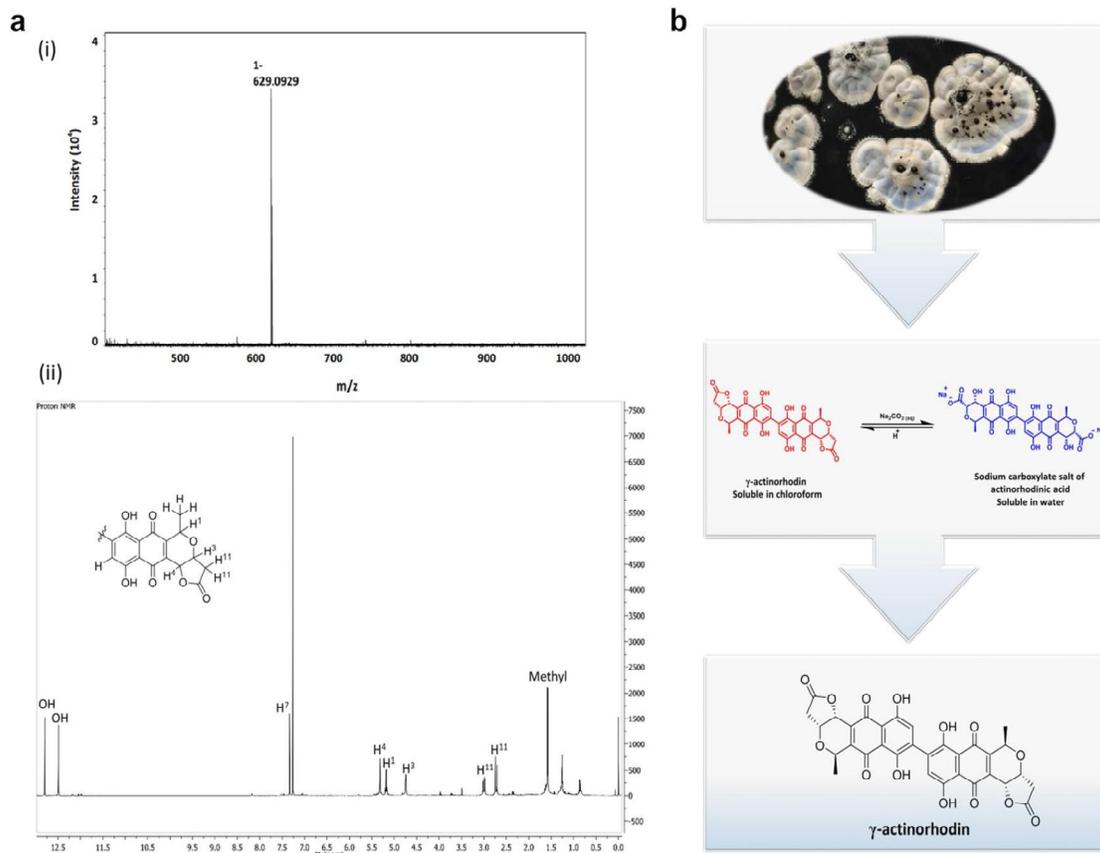


Fig. 1. Identification of γ -ACT as the predominant ACT species secreted by *S. coelicolor*L646 using LC-MS(ai) and ¹H NMR (aii), and overview of the γ -ACT purification strategy (b)

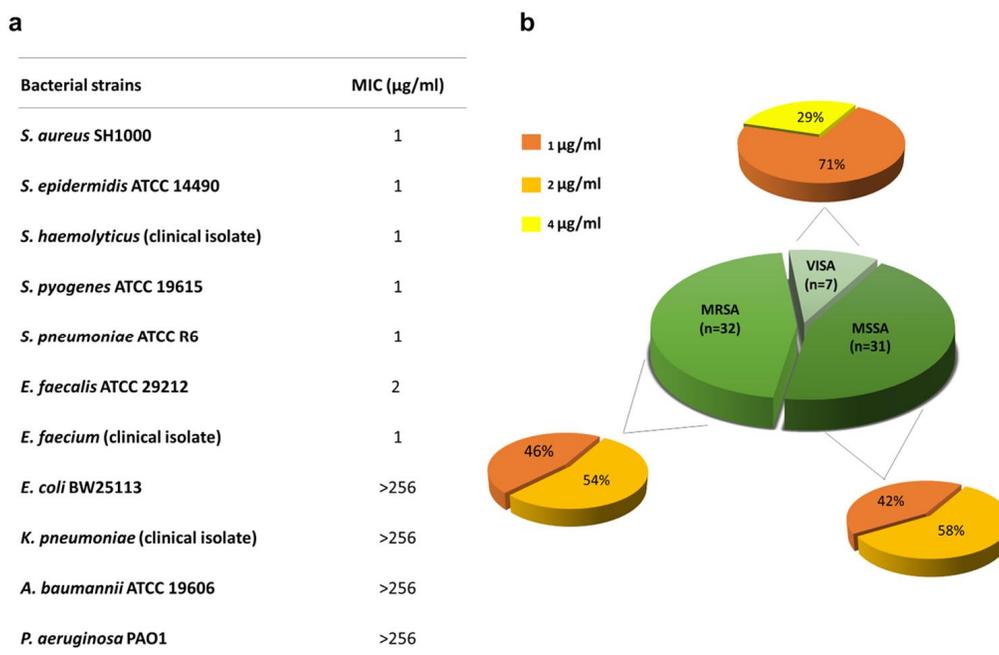


Fig. 2. *In vitro* antibacterial activity of γ -ACT

Hansen et al. [35] isolated two new cysteine-rich antibacterial peptides, turgencin A and turgencin B, from the Arctic marine colonial. Moreover, the peptides had contained methionine residues and the greatest active peptide is turgencin AMox1 with one oxidized methionine, showed anti microorganisms' activity with a minimum inhibitory concentration (MIC) against selected bacterial strains. For this reason, the peptide inhibited the growth of melanoma cancer and the human fibroblast cell line.

Díaz-Roa et al. (2019) identified and described a new *S. magellanica* AMP and their antimicrobial activity was evaluated and also, physicochemical characteristics estimated. Molecule sarconesin II showed activity against microorganisms, in addition, it was documented as a conserved domain of the ATP synthesis. The results concluded that the peptides would be utilizing in infections against microorganisms and a resource of compounds for fight multidrug-resistant bacteria.

AMPs were specified in necrophagous flies' salivary glands [36] and its significant components of larval excretions and secretions (ES). This indicated that digestive enzymes were produced by them are ingredients of ES [37]. These molecules' of action (MoA) was playing with ES ingredients, therefore gives the success

of larval treatment. More research has shown differentiate properties and estimate anti microorganisms' activity of blowfly larvae-derived molecules, involved AMPs [38].

AMPs had contained changed amino acid compounds, hydrophobic and have amphipathic properties [39]. These peptides interact with disease surface during the hydrophobic technique to begin death bacteria utilizing technique likes membrane permeabilization and rupture, involved reactive oxygen species (ROS) synthesis and due to cell death [40]. Variations essential series of the AMPs immediately effect of their technique of action, powder and selectivity against bacteria [41].

6. CLASSIFICATION OF SOME PRODUCTS PRODUCED FROM BACILLUS

The antimicrobial compounds of *Bacillus* sp. were utilized in agriculture for the inhibition of plant diseases, for the prompting of plant growth, and in the food industry (Wiyada, 2012).

Bioactive compounds of *Bacillus* sp. are split to two parts; (i) nonribosomal synthesis of cyclic lipopeptides (NRPS) and (ii) polyketides (PKS) (Table 1)., maybe caused varying properties.

Table 1. Major compounds from *Bacillus sp*

Compounds	Gene cluster involved	Bacillus species	Functions
Nonribosomal peptide synthetases (NRPS)			
Iturin	<i>itu, lpa</i>	B. subtilis, B. amyloliquefaciens	Antifungal, hemolytic activities
Fengycin	<i>fen, Pps</i>	B. subtilis, B. amyloliquefaciens	Anti-filamentous fungi
Surfactin	<i>srf, ycx, aat, sfp</i>	B. subtilis, B. amyloliquefaciens	Antiviral, antimycoplasma activities [42]
BacillomycinDa	<i>Bmy</i>	B. amyloliquefaciens	Antifungal
Bacillibactin	<i>Dhb</i>	B. subtilis, B. amyloliquefaciens	Iron transport system; siderophores
Putative peptide	<i>Nrs</i>	B. amyloliquefaciens	Siderophores (Herzner et al., 2008)
Bacilysin/anticapsin	<i>bac, ywf</i>	B. subtilis, B. amyloliquefaciens	Antimicrobial activity
ZwittermicinA	<i>Zwit</i>	B. cereus	Broad spectrum of antibacterial [43]
Nonribosomal peptide synthetases (NRPS)			
Subtilin	<i>Spa</i>	B. subtilis	Antimicrobial activity (lantibiotic)
SubtilosinA	<i>sbo, alb</i>	B. subtilis, B. amyloliquefaciens	Antimicrobial activity
TasA	<i>Tas</i>	B. subtilis	Antimicrobial activity
Sublancin	<i>sun, bdb</i>	B. subtilis	Antimicrobial activity (not lantibiotic)
Macrolactin	<i>Mln</i>	B. amyloliquefaciens	Anti-Gram-positive bacteria
Bacillaenea	<i>bae, pksX</i>	B. subtilis, B. amyloliquefaciens	Antibacterial activity
Difficidin	<i>Dif</i>	B. amyloliquefaciens	Antibacterial activity
Mersacidin	<i>Mrs</i>	B. amyloliquefaciens	Inhibit cell wall biosynthesis, anti-Gram-positive bacteria [44]

More strains *Bacillus sp* offer the capacity to secrete different bioactive ingredients of having great power commercial significance [45]. More lipopeptides (LPs) of *Bacillus* qualified thus away are divided into three major families: surfactins, iturins [46]. It could be caused their natural diversity LPs due to synthesize as a mixture of homologues and is forms varying in the amino acid composition of the peptide sequence [47]. Moreover, these biosurfactants stay active at maximum pH, salinity, and high temperatures. It was successfully lowering surface tension and also they are emulsifying and dispersing agents greatly utilized in more industrial sectors [48].

Members of the genus *Bacillus* are known to have multiple useful traits which help the plants immediately or indirectly during the acquisition of nutrients, overall improvement in growth by production of phytohormones, protection from pathogens and other abiotic stressors. This functionally versatile genus is one of the most

commercially exploited bacteria in the agrobiotechnology industry [49].

7. THE EFFECT OF ANTIBIOTIC FIND AND THE TROUBLE OF ANTIBIOTIC RESISTANCE

Antibiotics are the greatest effective forms of chemotherapy in medicine [50]. Antibiotics have given a share into excess in average life in advanced countries; indirectly in there to prevent disease utilize for help contagion next invasive surgeries [51]. Moreover, commonly utilize antibiotics in the therapy of infectious diseases has on condition the chosen compression required to be done to push the prevalence of antibiotic resistance (2015).

Elevating recurrence is multidrug-resistant (MDR) strains such as ESKAPE a bacterium, virus, or another microorganism that can cause disease [52]. A new report found that by 2050, the danger may be caused elevation of

antibacterial resistant infections [53]. After evaluation of resistance to the antibiotic colistin, the trouble has concentrated following a breaking other group of antibiotics against multidrug-resistant microorganisms infections [54]. The ability to be used antibiotics and colistin has driven to people information concerning being used the threat of bacterial resistance to antibiotics [55].

It is crucial to identify novel antibiotic classes which have shown antimicrobial activity against diseases which are resistant to antibiotics [28]. The pharmaceutical sector seems unable to respond to the threat of antibacterial resistance. The truth remains that best returns can be made in other areas of therapeutics [28]. As well as, bacterial resistance to antibiotics renders these compounds unsuccessful after a low duration of clinical utilize than with other pharmaceuticals [56].

8. STIMULATE BACTERIA IN LIQUID MEDIA TO PRODUCE MORE COMPOUNDS

Microorganisms are presents and predominately act reciprocally with all organisms sitting in the atmosphere, at some unspecified making a network of signals that shapes the basis for life on the Earth. These molecular signals, their production shape the absolutely necessary pre-requisite for deciphering inter-kingdom connections, characterized by or given to adaptation responses and systems biology [57].

Prodigiosin is a broad anti microorganisms' spectrum and encourages autolytic activity in the objective cells. Antibiotics are one bacteriocin produced by Gram-positive bacteria that can be bioengineered to increase their influences against a great number of bacterial strains and to become better their the state of being stable during the gastric transit that is by performance them protease-resistant [58] since it is common in causing disease, commensally, and food bacteria [59].

Other molecules are made by cytomegalovirus-infected cells: these compounds of viral origin (absolutely necessary proteins) can elevate virus dissemination, stability, and development of a disease by the act against host innate and characterized by immune responses. Conversely, more useful microbes, as Lactic Acid Bacteria (LAB) and Bifidobacteria can exert a modifying the immune system dominating

inflammation by means of proteins and non-proteinaceous compounds [60].

9. CONCLUSION

AMPs have gained importance as alternative agents for controlling spoilage and pathogenic micro-organisms. This article show reinforces?? the importance, isolated and purified of *Bacillus* species as a source of a variety of antimicrobial substances. Despite the fact that several AMPs are recognized by their low toxicity, an evaluation of the harmful effects on mammalian cells must be conducted before this peptide could be used in field experiments. The identification and characterization of novel AMPs and their potential use in the control of microbial infections are topics of greatest relevance.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

1. Sumi CD, Yang BW, Hahm YT. Antimicrobial peptides of the genus *Bacillus*: a new era for Antibiotics. *Can. J. Microbiol.* 2015;61:1–11.
2. Carvalho L, Remuzgo C, Perez KR, Machini MT. Hb40-61a: novel analogues help expanding the knowledge on chemistry, properties and candidacidal action of this bovine??-hemoglobin-derived peptide. *Biochim. Biophys. Acta Biomembr.* 2015;1848:3140–3149.
3. Bechinger B, Gorr SU. Antimicrobial peptides: mechanisms of action and resistance. *J. Dental Res.* 2017;96:254–260.
4. Brogden NK, Brogden KA. Will new generations of modified antimicrobial peptides improve their potential as pharmaceuticals? *Int. J. Antimicrob. Agents.* 2011;38(3):217–225.
5. Sang Y, Blecha F. Antimicrobial peptides and bacteriocins: alternatives to traditional

- antibiotics. *Anim. Health Res. Rev.* 2008;9(2):227–235.
6. Xie J, Zhang R, Shang C, Guo Y. Isolation and characterization of a bacteriocin produced by an isolated *Bacillus subtilis* LFB112 that exhibits antimicrobial activity against domestic animal pathogens. *Afr. J. Biotechnol.* 2009;8(20):5611–5619.
 7. Abriouel H, Franz CM, Omar NB, Gálvez A. Diversity and applications of *Bacillus* bacteriocins. *FEMS Microbiol. Rev.* 2011; 35(1):201–232.
 8. Pletzer D, Mansour SC, Hancock REW. Synergy between conventional antibiotics and anti-biofilm peptides in a murine, sub-cutaneous abscess model caused by recalcitrant ESKAPE pathogens. *PLoS Pathog.* 2018;14:e1007084. DOI: 10.1371/journal.ppat.1007084
 9. Sabaté DC, Audisio MC. Inhibitory activity of surfactin, produced by different *Bacillus subtilis* subsp. *subtilis* strains, against *Listeria monocytogenes* sensitive and bacteriocin-resistant strains. *Microbiol. Res.* 2013;168(3):125–129.
 10. Marx R, Stein T, Entian KD, Glaser SJ. Structure of the *Bacillus subtilis* peptide antibiotic subtilosin A determined by 1H-NMR and matrix assisted laser desorption/ionization time-of-flight mass spectrometry. *J. Protein Chem.* 2001; 20(6):501–506.
 11. Li G, Liu B, Shang Y, Yu Z, Zhang R. Novel activity evaluation and subsequent partial purification of antimicrobial peptides produced by *Bacillus subtilis* LFB112. *Ann. Microbiol.* 2012;62(2):667–674.
 12. Benitez LB, Velho RV, Lisboa MP, Medina LF, Brandelli A. Isolation and characterization of antifungal peptides produced by *Bacillus amyloliquefaciens* LBM5006. *J. Microbiol.* 2010;48(6):791–797.
 13. Xu MH, Rong YJ, Zhao MX, Song B, Chi, ZM. Antibacterial activity of the lipopeptides produced by *Bacillus amyloliquefaciens* M1 against multidrug-resistant *Vibrio* spp. isolated from diseased marine animals. *Appl. Microbiol. Biotechnol.* 2014;98(1):127–136.
 14. Ayed HB, Hmidet N, Béchet M, Chollet M, Chataigné G, Leclère V. Identification and biochemical characteristics of lipopeptides from *Bacillus mojavensis* A21. *Process Biochem.* 2014;49(10):1699–1707.
 15. Chopra L, Singh G, Choudhury V, Shao DK. Sonorensin: an antimicrobial peptide, belonging to the heterocycloanthracin subfamily of bacteriocins, from a new marine isolate, *Bacillus sonorensis* MT93. *Appl. Environ. Microbiol.* 2014;80(10):2981–2990.
 16. Dimkic´ I, Žickovic´ S, Beric´ T, Ivanovic´ Ž, Gavrilovic´ V, Stankovic´ S, Fira D. Characterization and evaluation of two *Bacillus* strains, SS-12.6 and SS-13.1, as potential agents for the control of phytopathogenic bacteria and fungi. *Biol. Control.* 2013;65(3):312–331.
 17. Al-Thubiani ASA, Maher YA, Fathi A, Abourehab MAS, Alarjah M, Khan MSA. And Al- Ghamdi SB. Identification and characterization of a novel antimicrobial peptide compound produced by *Bacillus megaterium* strain isolated from oral Microflora, *Saudi Pharmaceutical Journal.* 2018;26:1089–1097.
 18. Cutting SM. *Bacillus* probiotics. *Food Microbiol.* 2011;28(2):214–220.
 19. Haddar HO, Aziz GM, Al-Gelawi MH. Optimization of bacitracin production by *Bacillus licheniformis* B5. *Pak. J. Biol. Sci.* 2007;10(6):972–976. DOI:10.3923/pjbs.2007.972.976
 20. Van Kuijk S, Noll KS, Chikindas ML. The species-specific mode of action of the antimicrobial peptide subtilosin against *Listeria monocytogenes* Scott A. *Lett. Appl. Microbiol.* 2011;54(1):52–58.
 21. Zavascki AP, Goldani LZ, Li J, Nation RL. Polymyxin B for the treatment of multidrug-resistant pathogens: A critical review. *J. Antimicrob. Chemother.* 2007;60:1206–1215.
 22. Choi SK, Park SY, Kim R, Kim SB, Lee CH, Kim JF, Park SH. Identification of a polymyxin synthetase gene cluster of *Paenibacillus polymyxa* and heterologous expression of the gene in *Bacillus subtilis*. *J. Bacteriol.* 2009;191(10):3350–3358.
 23. Wang, Yof microcapsules containing antimicrobial lipopeptide from *Bacillus amyloliquefaciens* ES-2 by spray drying. *Food Sci. Technol.* 2014;56(2):502–507.
 24. Jung WJ, Mabood F, Souleimanov A, Zhou X, Jaoua S, Kamoun F, Smith DL. Stability and antibacterial activity of bacteriocins produced by *Bacillus thuringiensis* and *Bacillus thuringiensis* ssp. *kurstaki*. *J. Microbiol. Biotechnol.* 2008;18(11):1836–1840.
 25. Halimi B, Dortu C, Arguelles-Arias A, Thonart P, Joris B, Fickers P. Antilisterial

- activity on poultry meat of amylolysin, a bacteriocin from *Bacillus amyloliquefaciens* GA1. *Probiotics Antimicrob. Proteins*. 2010;2(2):120–125.
26. Guo Q, Dong W, Li S, Lu X, Wang P, Zhang X. Fengycin produced by *Bacillus subtilis* NCD-2 plays a major role in biocontrol of cotton seedling damping-off disease. *Microbiol. Res.* 2014;169(7–8):533–540.
 27. Nass NM, Farooque S, Hind C, Wand ME, Randall CP, Sutton JM, Seipke RF, Rayner CM, O'Neill AJ. Revisiting unexploited antibiotics in search of new antibacterial drug candidates: the case of γ -actinorhodin. *SCientificREPORts* | 2017; 7:17419,1-11.
 28. Silver LL. Challenges of antibacterial discovery. *Clin Microbiol Rev.* 2011;24:71–109.
 29. Nishiyama T, Hashimoto Y, Kusakabe H, Kumano T, Kobayashi M. Natural low-molecular mass organic compounds with oxidase activity as organocatalysts. *Proc Natl Acad Sci USA.* 2014;111:17152–17157.
 30. Randall CP, Mariner KR, Chopra I, O'Neill AJ. The target of daptomycin is absent from *Escherichia coli* and other Gram-negative pathogens. *Antimicrob Agents Chemother.* 2013;57:637–639.
 31. Kumar P, Kizhakkedathu JN, Straus SK. Antimicrobial Peptides: diversity, mechanism of action and strategies to improve the activity and biocompatibility in vivo. *Biomolecules.* 2018;8.
 32. Elhag O, Zhou D, Song Q, Soomro AA, Cai M, Zheng L, Yu Z, Zhang J. Screening, Expression, Purification and Functional Characterization of Novel Antimicrobial Peptide Genes from *Hermetia illucens* (L.). *PLoS ONE*, 2017;12:e0169582.
 33. Romoli O, Saviane A, Bozzato A, D'Antona P, Tettamanti G, Squartini A, Cappellozza S, Sandrelli F. Differential sensitivity to infections and antimicrobial peptide-mediated immune response in four silkworm strains with different geographical origin. *Sci. Rep.* 2017;7:1048.
 34. Rifflet A, Gavalda S, Téné N, Orivel J, Leprince J, Guilhaudis L, Génin E, Vétillard A, Treilhou M. Identification and characterization of a novel antimicrobial peptide from the venom of the ant *Tetramorium bicarinatum*. *Peptides.* 2012; 38:363–370.
 35. Hansen IK, Isaksson J, Poth AJ, Hansen K, Andersen AJC, Richard CM, Blencke H-M, Stensvåg K, Craik DJ, Haug T. Isolation and Characterization of Antimicrobial Peptides with Unusual Disulfide Connectivity from the Colonial Ascidian *Synoicum turgens*. *Mar. Drugs.* 2020;18:51.
 36. Valachova I, Bohova J, Palosova Z, Takac P, Kozanek M, Majtan J. Expression of lucifensin in *Luciliasericata* medicinal maggots in infected environments. *Cell Tissue Res.* 2013;353:165–171.
 37. Pinilla YT, Patarroyo MA, Velandia ML, Segura NA, Bello FJ. The effects of *Sarconesiopsis magellanica* larvae (Diptera: Calliphoridae) excretions and secretions on fibroblasts. *Acta Trop.* 2015;142:26–33.
 38. Yakovlev AY, Nesin AP, Simonenko NP, Gordya NA, Tulin DV, Kruglikova AA, Chernysh SI. Fat body and hemocyte contribution to the antimicrobial peptide synthesis in *Calliphoravicina* R.-D. (Diptera: Calliphoridae) larvae. *In Vitro Cell. Dev. Biol. Anim.* 2017;53:33–42.
 39. Mishra AK, Choi J, Moon E, Baek KH. Tryptophan-Rich and Proline-Rich Antimicrobial Peptides. *Molecules.* 2018;23.
 40. Yi HY, Chowdhury M, Huang YD, Yu XQ. Insect antimicrobial peptides and their applications. *Appl. Microbiol. Biotechnol.* 2014;98:5807–5822.
 41. Manzo G, Ferguson PM, Gustilo VB, Hind CK, Cliford M, Bui TT, Drake AF, Atkinson RA, Sutton JM, Batoni G. Minor sequence modifications in temporin B cause drastic changes in antibacterial potency and selectivity by fundamentally altering membrane activity. *Sci. Rep.* 2019;9:1385.
 42. Vollenbroich D, Pauli G, Ozel M, Vater J. Antimycoplasma properties and application in cell culture of surfactin, a lipopeptide antibiotic from *Bacillus subtilis*. *Appl. Environ. Microbiol.* 1997;63:44–49.
 43. He H, Silo-Suh LA, Clardy J, Handelsman J. Zwittermicin A, an antifungal and plant protection agent from *Bacillus cereus*. *Tetrahedron Lett.* 1994;35:2499–2502.
 44. Brötz H, Bierbaum G, Leopold K., Reynolds PE, Sahl HG. The lantibiotic mersacidin inhibits peptidoglycan synthesis by targeting lipid II. *Antimicrob. Agents Chemother.* 1998;42:154–160.
 45. Jacques P. Surfactin and other lipopeptides from *Bacillus* spp. In *Biosurfactants: From Genes to*

- Applications ed. Soberon-Chavez, G. Berlin Heidelberg: Springer, Microbiology Monographs. 2011;57–93:20.
46. Mongkolthananuk W. Classification of Bacillus beneficial substances related to plants, humans and animals. J Microbiol Biotechnol. 2012;22:1597–1604.
 47. Pecci Y, Rivardo F, Martinotti MG, Allegrone G. LC/ESI-MS/MS characterization of lipopeptide bio-surfactants produced by the Bacillus licheniformis V9T14 strain. J Mass Spectrom 2010;45:772–778.
 48. Mulligan CN, Sharma SK, Mudhoo A. Biosurfactants. Research Trends and Applications. Boca Raton, London, New York: CRC Press Taylor & Francis Group; 2014.
 49. Saxena AK, Kumar M, Chakdar H, Anuroopa N, Bagyaraj DJ. Bacillus species in soil as a natural resource for plant health and nutrition, Journal of Applied Microbiology. 2019;128:1583–1594.
 50. Aminov RI. A brief history of the antibiotic era: lessons learned and challenges for the future. Frontiers in Microbiology. 2010;1.
 51. Ventola CL. The antibiotic resistance crisis: part 1: causes and threats. P T. 2015; 40(4):277-83.
 52. CDC. Antibiotic resistance threats in the United States. (1560-7917 (Electronic) 1025-496X (Linking)). Atlanta: Centers for Disease Control and Prevention; 2013.
 53. O'Neill J. Tackling drug-resistant infections globally: final report and recommendations. Review on antimicrobial resistance; 2016.
 54. Zeng KJ, Dol YH, Patil S, Huang X, Tian GB. Emergence of the plasmid-mediated mcr-1 gene in colistin-resistant Enterobacter aerogenes and Enterobacter cloacae. Antimicrobial Agents and Chemotherapy. 2016;60(6):3862-3863.
 55. Chen LR, Todd J, Kiehlauch M. Walters, Kallen A. Notes from the field: Pan-resistant New Delhi Metallo-beta-lactamase-producing Klebsiella pneumoniae - Washoe County, Nevada, 2016. MMWR Morb Mortal Wkly Rep. 2017;66(1):33.
 56. O'oi N, O'Neill AJ. Revisiting unexploited antibiotics in search of new antibacterial drug candidates: the case of MSD-819 (6-chloro-2-quinoxalinecarboxylic acid 1,4-dioxide). J Antibiot (Tokyo). 2017;70(3): 317-319.
 57. Cani PD, Knauf C. How gut microbes talk to organs: the role of endocrine and nervous routes. Mol. Metab. 2016;5:743–752.
 58. WHO. World Health Organization. Antimicrobial resistance: 2014 Global Report on Surveillance. Geneva: World Health Organization; 2014.
 59. Laxminarayan R, Duse A, Wattal C, Zaidi AKM, Wertheim HFL, Sumpradit N. Antibiotic resistance-the need for global solutions. Lancet Infect. Dis. 2013;13: 1057–1098.
 60. Pessione E. Lactic acid bacteria contribution to gut microbiota complexity: lights and shadows. Front. Cell. Infect. Microbiol. 2012;2:86.

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